

ORIGINAL ARTICLE

Extremely High Mortality Rates in Patients with Carbapenem-resistant, Hypermucoviscous *Klebsiella pneumoniae* Blood Stream Infections

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Abstract

Background: Infections caused by carbapenem resistant *K. pneumoniae* are increasing and associated with high mortality rates. There are increasing reports of hypermucoviscous/ hypervirulent *K. pneumoniae* isolated from various sources. However, there is limited data on the prevalence of hypermucoviscous strains among carbapenem-resistant *K. pneumoniae* from invasive infections in India and its association with mortality. *rmpA*, *rmpA2* and *magA* genes are associated with these hypervirulent strains. In this study, we investigate the prevalence of hypermucoviscous strains amongst carbapenem resistant *K. pneumoniae* isolated from blood culture. Association of mortality rate with meropenem minimum inhibitory concentration and hypermucoviscous strains are determined.

Methods: 86 non-repetitive carbapenem resistant *K. pneumoniae* isolated from bacteremia underwent E-test for meropenem minimum inhibitory concentration (MIC) determination and PCR for detection of carbapenamase genes. String test, PCR for *rmpA*, *rmpA2* and *magA* were performed for characterisation of hypervirulent strains.

Results: 31.3% of the 86 isolates displayed hypermucoviscous phenotype as indicated by a positive string test. Among the two genotypic markers, 7% were positive for *rmpA2* and all were negative for *rmpA* and *magA*. 74.1% and 67.9% mortality were seen among string test positives and isolates meropenem MIC of $\geq 16\mu\text{g/ml}$ respectively (p 0.036 and 0.008 respectively). Isolates with both string positivity and meropenem MIC of $\geq 16\mu\text{g/ml}$ had a very high mortality rate of 84.2%.

Conclusion: String test, aids prediction of disease severity, and is independently associated with increased mortality in invasive carbapenem resistant *K. pneumoniae* health care-acquired infections. High meropenem MIC is a significant risk factor for mortality. Combination of string positive carbapenem resistant hypermucoviscous *K. pneumoniae* resulted in mortality rate of 84.2%. It is important to monitor prevalence of carbapenem resistant hypermucoviscous/ hypervirulent *K. pneumoniae* among invasive isolates especially in a setting with high resistance rates as combination of increased virulence and decreased susceptibility to antimicrobials results in worse outcomes.

Increased and poorly regulated antibiotic use has resulted in a rise in incidence of carbapenem-resistant *K. pneumoniae* (CRKp). These infections have a very high mortality rate of 30 to 44%.⁴⁻⁶

Recent years have also brought a rise in incidence of a hypermucoviscous/ hypervirulent variant of *K. pneumoniae* (hvKp) associated with liver abscess.⁷ The string test is a phenotypic marker used to screen for these strains. The genes *rmpA* and *rmpA2* (both regulators of mucoid phenotype) and *magA* (mucoviscosity associated gene) are associated with string test positivity and are sometimes used as genomic markers of hypervirulence.⁷ Other virulence genes that are expressed more frequently in hypervirulent *K. pneumoniae* than the classical *K. pneumoniae* include siderophores such as *iutA* (aerobactin), *ybtS* (yersiniabactin), *entB* (enterobactin), *kfu* (mediating ferric iron uptake); *allS* (allantoin metabolism) and *mrkD* (type3 fimbriae mediating attachment to extracellular matrix).⁸

Unfortunately, there is no internationally agreed definition for hvKp strain and this hinders surveillance and literature review. Some authors define hypervirulence by clinical syndrome alone including patients with liver abscess, metastatic infection and high mortality rate caused by *K. pneumoniae*. Others define hypervirulence phenotypically by string test positivity or genotypically by *rmpA* or *rmpA2* positivity. Further authors use mouse lethality studies.⁹⁻¹³

Background

Klebsiella pneumoniae is a common pathogen and causes a wide range of infections including pneumonia, urinary tract infection, intra-abdominal infection and wound infection.¹ Infections with *K. pneumoniae* are usually hospital-acquired and occur

primarily in patients with impaired host defences² including solid tumours, haematological malignancies, liver cirrhosis and diabetes mellitus.³

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Received: 07.06.2017; Accepted: 12.09.2018

Table 1: List of primers used in the study

Gene	Forward primer (5'-3')	Reverse primer (5'-3')	Reference
<i>bla</i> _{IMP}	GGAATAGAGTGGCTTAAYCTC	GGTTTAAAYAAAACAACCACC	16
<i>bla</i> _{VIM}	GATGGTGTGGTTCGCATA	CGAATGCGCAGCACCAG	19
<i>bla</i> _{NDM}	CACCTCATGTTGAATTCGCC	CTCTGTCACATCGAAATCGC	16
<i>bla</i> _{OXA-48-like}	TATATTGCATTAAGCAAGGG	CACACAAATACGCGCTAACC	16
<i>bla</i> _{KPC}	TGTCACTGTATCGCCGTC	CTCAGTGTCTACAGAAAACC	18
<i>bla</i> _{SPM}	AAAATCTGGGTACGCAAACG	ACATTATCCGCTGGAACAGG	17
<i>rmpA</i>	TGTTAACATGCAAGGAAATG	ATTGCAGCACTGCTTGTT	This study
<i>rmpA2</i>	CTTTATGTGCAATAAGGATGT	CCTCCTGGAGAGTAAGCATT	10
<i>magA</i>	GGTGCTCTTACATCATTCG	GCAATGGCCATTTGCGTTAG	10

There have been no reports on the prevalence of hypermucoviscous variants among CRKp causing bacteremia in India. This study aims to characterize carbapenem-resistant hypermucoviscous strains isolated from bacteremia. We investigate the phenotypic and genotypic prevalence of the hypermucoviscous together with meropenem MIC and carbapenemase genes. The association of string test and meropenem MIC with patient mortality was determined.

Methods

Study population

The study was performed at the department of Clinical Microbiology, Christian Medical College, Vellore, India. Cases of bacteremia from 2014 and 2015 were considered and 86 non-repetitive, first positive blood culture isolated from patients that grew carbapenem resistant *K. pneumoniae* were included in the study.

Phenotypic characterisation of bacterial isolates:

Isolation and identification of the *K. pneumoniae* isolates were performed using standard biochemical reactions. The antimicrobial susceptibility testing for the first and second line antimicrobials was performed using Kirby Bauer disc diffusion as recommended by CLSI (Clinical and Laboratory Standards Institute) and the results were interpreted according to CLSI guidelines. 86 carbapenem resistant isolates were chosen. Further, these were tested for meropenem by E-test (Biomérieux) to determine the minimum inhibitory concentration (MIC). *E. coli* ATCC 25922 was used as control strain for antimicrobial susceptibility testing.

String test was performed for the detection of hypermucoviscous *K. pneumoniae* and was considered positive when a viscous string of >5mm was

produced when the colonies were stretched with an inoculation loop.¹⁵

Molecular characterization of bacterial isolates

Bacterial genomic DNA was isolated by boiling the bacterial suspension at 100°C for 15 minutes and the supernatant was collected after centrifugation and used for molecular assays. Molecular characterization of bacterial isolates was carried out using conventional multiplex PCR for the detection of carbapenemase genes which included *bla*_{IMP}, *bla*_{VIM}, *bla*_{NDM}, *bla*_{SPM}, *bla*_{OXA 48-like} and *bla*_{KPC} as described by Poirel *et al.*, 2011, Dallene *et al.*, 2010, Yigit *et al.*, 2001 and Ellington *et al.*, 2007.¹⁴⁻¹⁷ For the detection of hypervirulent *K.pneumoniae*, *rmpA*, *rmpA2* and *magA* were detected. PCR for *rmpA2* and *magA* were performed as described by Brisse *et al.*, 2009.¹⁰ *magA* also corresponds to the capsular type1. The primers used are as mentioned in Table 1.

Multi-locus Sequence Typing

Ten representative isolates were sequenced and typed using the protocol mentioned by Diancourt *et al.*, 2005.¹⁸ Four string test positive isolates and six string test negative isolates were typed. The sequence type was determined using the database maintained by Pasteur Institute at <http://bigsd.bpasteur.fr/klebsiella/klebsiella.html>

Clinical details

Clinical details of the patients, including age, sex, co-morbidities, source of infection and clinical outcome, were collected retrospectively from electronic medical records at Christian Medical College, Vellore.

Statistical analysis

Results were analysed for association between 30-day mortality with meropenem MIC, string test positivity and carbapenemase genes using Chi square test and logistic regression with 95% CI. The variables associated

Table 2: Demographic details of patients with carbapenem resistant *K. pneumoniae* (n=86)

	Numbers (%)
Sex	
Male	56 (65)
Female	30 (35)
Age	
Range: 0.002 to 80 yrs	-
Median: 37.5yrs	
Immune status	
Immunocompromised	45 (52)
Other co-morbidities	28 (33)
No comorbidities	13 (15)
Nature of infection	
Health-care Acquired	79 (92)
Community Acquired	5 (6)
Unclear	2 (2)
Source of infection	
Neutropenic sepsis	29 (33)
Abdominal sepsis	20 (23)
Respiratory sepsis	15 (16)
Unknown	9 (10)
Central/peripheral line	6 (7)
Neonatal sepsis	5 (6)
Skin and soft tissue infection/ bone/joint	4 (5)

with risk factors for mortality were considered significant if the *p* value was <0.05. For analysis of meropenem MIC, isolates were stratified into four groups based on meropenem MIC. The first group comprised of susceptible isolates with MIC ≤1µg/ml (n=11), the second of intermediate susceptible isolates with MIC 2 µg/ml (n=11), and the third of resistant isolates with MIC between ≥4 µg/ml to ≤16 µg/ml (n=11). Since a significant proportion of the isolates in the study had MIC of ≥16 µg/ml (n=53), a fourth group considered these isolates that were highly resistant to meropenem.

Results

CRKp isolates from 86 patients obtained during 2015 and 2016 were included in this study. 65% of the patients included in the study were male with a median age of 37.5 years. 52% were immunocompromised. 92% of infections were associated with healthcare. Further clinical details can be found in Table 2.

Twenty-seven (31.3%) isolates were string test positive and 59 (68.6%) were negative. Only three string positive and three string negative isolates contained the *rmpA2* gene. *rmpA* was absent in all the isolates. None of the isolates belonged to K1 capsular type since

Table 3: Distribution of carbapenamase genes among the *K pneumoniae* resistant to carbapenems

Carbapenamase genes	No. of positives (n=86)	p value	String test positives n=27	String test negatives n=59
<i>bla</i> _{NDM}	31 (36%)	0.203	11 (41%)	20 (34%)
<i>bla</i> _{OXA48 like}	23 (27%)	0.123	6 (22%)	17 (29%)
<i>bla</i> _{NDM and OXA48-like}	23 (27%)	0.967	10 (37%)	13 (22%)
<i>bla</i> _{NDM, OXA48 like and VIM}	3 (3%)	0.134	0	3 (5%)
<i>bla</i> _{NDM and VIM}	3 (3%)		0	3 (5%)
<i>bla</i> _{VIM and OXA48 Like}	3 (3%)		0	3 (5%)

they lacked *magA* gene. The results of the multiplex PCR for carbapenamase genes are tabulated in Table 3. *bla*_{NDM} was the most common gene expressed by the isolates followed by *bla*_{OXA48 like}, but there was no difference in the distribution among string test positives and negatives.

30 day mortality was 55.9% (n=49). There was no association between mortality and age, sex, co-morbidity, immunosuppression or carbapenamase gene. However, there was a positive association between mortality rate and meropenem MIC (Figure 1). Meropenem MIC ≥ 16 $\mu\text{g/ml}$ was a significant and independent risk factor for mortality (p 0.008, OR 9.5). String test positivity was also independently associated with mortality (p 0.036, OR 2.23).

Among the ten isolates that were typed, six were negative by string test and four were positive. Among the isolates that were string test negative, three belonged to ST14 and ST231 each. Among the four isolates positive for string test, two isolates belonged to ST231 and one each to ST11 and ST43. However, both the string test positive isolates belonging to ST231 lacked *rmpA* and *rmpA2* genes.

Discussion

CRKp are a common cause of nosocomial infections and have previously been associated with a high 30 day mortality rate of 42%.^{19,20} Ben-David *et al.*, found that carbapenam resistance is an independent risk factor for mortality in blood stream infections caused by *K. pneumoniae*.²¹ In this study 92% of the CRKp infections were health-care associated and the overall mortality was higher than previously described at 55.9%. In this study, we describe increasing mortality rate with increasing meropenem MICs. As mentioned in Figure 1, the mortality in patients with meropenem MIC ≤ 1 $\mu\text{g/ml}$ is 20% but is double this for patients

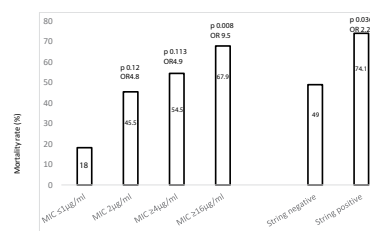
with MIC of 2 $\mu\text{g/ml}$. Mortality rates in patients with MIC ≥ 4 $\mu\text{g/ml}$ and ≥ 16 $\mu\text{g/ml}$ are 54.5% and 67.9% respectively.

In India, carbapenam resistance is mostly due to the NDM and OXA enzymes^{22,23} as seen in this study while in Europe KPC is the major carbapenamase encoded by the bacteria. However, the type of carbapenamase gene did not correlate with meropenem MIC or outcome ($p > 0.05$) as shown in Table 3. Sixty two percent of the isolates had a MIC of > 32 $\mu\text{g/ml}$ for meropenem and this was found to be an independent risk factor for mortality (p 0.008).

Immunosuppression (p 0.049) and high meropenem MICs (p 0.03) have found to be risk factors for mortality in CRKp infections in other studies.²⁴ Although we found that meropenem MIC ≥ 16 $\mu\text{g/ml}$ (p 0.008) was a significant risk factor, immunosuppression (p 0.533) was not. Nature and source of infection as mentioned in Table 2 were not risk factors mortality.

hvKp usually causes community acquired sepsis with a high mortality (55%)²⁵ and is almost always susceptible to beta-lactam antibiotics. There is limited data on hospital acquired CR-hvKp infections which we describe in this study. Very few cases of CR-hvKp has been described³³; one such being from China where a single isolate was obtained from tracheal secretion and was pan drug resistant.²⁶ The other report by Zhang *et al.*, 2015, describe five CR-hvKp causing pneumonia, septicaemia, abdominal infection and sepsis.²⁷

Laboratory screening method for detection of hvKp is by the string test which has a cut off of ≥ 5 mm for the formation of a viscous string. A modification with a cut-off of ≥ 10 mm has been proposed by Lee *et al.*, 2010 to improve the specificity.²⁸ Tan *et al.*, found that the test sensitivity (90.5%) and specificity (63%-66%) remained

**Fig. 1: Logistic regression for meropenem MIC and string positivity with mortality**

the same with both the cut-off values.²⁹ Although *rmpA*, *rmpA2* and *magA* are associated with positive string test, in this study we found only three *rmpA2* positives among the 27 string test positive. Isolates which are phenotypically hypermucoviscous but lack the *rmpA* and *rmpA2* genes could be due to the presence of other genes producing the expression of the phenotype. Conversely, isolates with *rmpA* and *rmpA2* genes which lack the phenotype may be due to mutations leading to truncated proteins³⁰ or, as previously published in ESBL producers, resistance genes which are expressed over virulence genes.³⁰ Other possible reasons could be the lack of another positive regulator for *rmpA* and the presence of negative regulators at post-transcriptional level. Further work investigating discrepancies between phenotypic and genotypic markers for hvKp is needed to allow the development of a formal definition of hypervirulent *K. pneumoniae*.

The community acquired hypervirulent *K. pneumoniae* syndrome is principally caused by the CC23 strains. In this study, MLST was performed only for 10 isolates and were found to belong to ST11, ST14, ST43 and ST231. These clonal types have previously been reported in carbapenam resistant *K. pneumoniae*; ST14 has been reported in NDM-1 producing isolate from India³¹ and ST231 in NDM *K. pneumoniae* in Spain.³² ST11 has also been reported among hypervirulent carbapenam resistant *K. pneumoniae* from China³⁴ coding for KPC carbapenamase unlike the isolate in the present study which produced NDM. Since the isolates did not predominantly belong to a single clonal type, surveillance of antimicrobial resistance and virulence is needed in order to prevent new clones from acquiring resistance and virulence.

Conclusion

String test is a quick phenotypic test for detection of hvKp which aids prediction of disease severity and is independently associated with increased mortality in invasive CRKp health care-acquired infections. High meropenem MIC is a significant risk factor for mortality. The combination of CRKp and string positive hvKp resulted in a mortality rate of 84.2%. It is important to monitor the prevalence of CR-hvKp among invasive isolates as they pose a public health threat in the treatment and management of infections. The combination of increased virulence and decreased susceptibility to antimicrobials is very challenging to treat and hence results in worse outcomes. Further work to define the hypervirulent strains with confirmatory markers are necessary since these strains are now seen globally. This will enable easier diagnosis and better management of the patients. Resistance and virulence are not restricted to a single clonal type in the present study reflecting on the diversity of clones present.

Ethics approval and consent to participate

Not applicable since this is a retrospective study in which the isolates were used without the patient identifier.

Competing interests

The authors declare that they have no competing interests.

Acknowledgements

We would like to thank Mr. Georgekutty Mathew, Department of Biostatistics, Christian Medical College, Vellore, India, for his assistance in statistical analysis.

References

- Podschun R, Ullmann U. *Klebsiella* spp. as nosocomial pathogens: epidemiology, taxonomy, typing methods, and pathogenicity factors. *Clin Microbiol Rev* 1998; 11:589–603.
- Moon HW, Ko YJ, Park S, Hur M, Yun YM. Analysis of community- and hospital acquired bacteraemia during a recent 5-year period. *J Med Microbiol* 2014; 63:421–6.
- Zammit SC, Azzopardi N, Sant J. Mortality risk score for *Klebsiella pneumoniae* bacteraemia. *European Journal of Internal Medicine* 2014; 25:571–6.
- Schwaber M, Klarfeld-Lidji S, Navon-Venezia S, Schwartz D, Levavt A, Carmeli Y. Predictors of carbapenem-resistant *Klebsiella pneumoniae* acquisition among hospitalized adults and effect of acquisition on mortality. *Antimicrob Agents Chemother* 2008; 52:1028–1033.
- Patel G, Huprikar S, Factor SH, Jenkins SG, Calfee P. Outcomes of carbapenem-resistant *Klebsiella pneumoniae* infection and impact of antimicrobial and adjunctive therapies. *Infect Control Hosp Epidemiol* 2008; 29:1099–1106.
- Falagas ME, Rafailidis PI, Kofteridis D, Vrtizili S, Chelvatoglou FC, Papaioannou V, et al. Risk factors of carbapenem-resistant *Klebsiella pneumoniae* infections: a matched case-control study. *Journal of Antimicrobial Chemotherapy* 2007; 60:1124–30.
- Shon AS, Bajwa RP, Russo TA. Hypervirulent (hypermucoviscous) *Klebsiella pneumoniae*: a new and dangerous breed. *Virulence* 2013; 4:107–18.
- Compain F, Babosan A, Brisse S, Genel N, Audo J, Ailloud F, et al. Multiplex PCR for detection of seven virulence factors and K1/K2 capsular serotypes of *Klebsiella pneumoniae*. *Journal of Clinical Microbiology* 2014; 52:4377–80.
- Wu KM, Li LH, Yan JJ, Tsao N, Liao TL, Tsai HC, et al. Genome sequencing and comparative analysis of *Klebsiella pneumoniae* NTUH-K2044, a strain causing liver abscess and meningitis. *Journal of Bacteriology* 2009; 191:4492–501.
- Brisse S, Fevre C, Passet V, Issenuth-Jeanjean S, Tournebise R, Diancourt L, et al. Virulent clones of *Klebsiella pneumoniae*: identification and evolutionary scenario based on genomic and phenotypic characterization. *PLoS one* 2009; 4:e4982.
- Russo TA, Shon AS, Beanan JM, Olson R, MacDonald U, Pomakov AO, et al. Hypervirulent *K. pneumoniae* secretes more and more active iron-acquisition molecules than “classical” *K. pneumoniae* thereby enhancing its virulence. *PLoS One* 2011; 6:e26734.
- Lin YC, Lu MC, Tang HL, Liu HC, Chen CH, Liu KS, et al. Assessment of hypermucoviscosity as a virulence factor for experimental *Klebsiella pneumoniae* infections: comparative virulence analysis with hypermucoviscosity-negative strain. *BMC Microbiology* 2011; 11:1.
- Fang CT, Chuang YP, Shun CT, Chang SC, Wang JT. A novel virulence gene in *Klebsiella pneumoniae* strains causing primary liver abscess and septic metastatic complications. *J Exp Med* 2004; 199:697–705.
- Poirel L, Walsh TR, Cuvillier V, Nordmann P. Multiplex PCR for detection of acquired carbapenemase genes. *Diagn Microbiol Infect Dis* 2011; 70:119–23.
- Ellington MJ, Kistler J, Livermore DM, Woodford N. Multiplex PCR for rapid detection of genes encoding acquired metallo-β-lactamases. *Journal of Antimicrobial Chemotherapy* 2007; 59:321–2.
- Yigit H, Queenan AM, Anderson GJ, Domenech-Sanchez A, Biddle JW, Steward CD, et al. Novel carbapenem-hydrolyzing beta-lactamase, KPC-1, from a carbapenem-resistant strain of *Klebsiella pneumoniae*. *Antimicrob Agents Chemother* 2001; 45:1151–61.
- Dallenne C, Da Costa A, Decré D, Favier C, Arlet G. Development of a set of multiplex PCR assays for the detection of genes encoding important beta-lactamases in Enterobacteriaceae. *J Antimicrob Chemother* 2010; 65:490–5.
- Diancourt L, Passet V, Verhoef J, Grimont PA, Brisse S. Multilocus sequence typing of *Klebsiella pneumoniae* nosocomial isolates. *Journal of Clinical Microbiology* 2005; 43:4178–82.
- Tumbarello M, Viale P, Viscoli C, Trearicchi EM, Tumietto F, Marchese A, et al. Predictors of mortality in bloodstream infections caused by *Klebsiella pneumoniae* carbapenemase-producing *K. pneumoniae*: importance of combination therapy. *Clinical Infectious Diseases* 2012; 55:943–50.
- Nguyen M, Eschenauer GA, Bryan M, O’Neil K, Furuya EY, Della-Latta P, et al. Carbapenem-resistant *Klebsiella pneumoniae* bacteremia: factors correlated with clinical and microbiologic outcomes. *Diagnostic Microbiology and Infectious Disease* 2010; 67:180–4.
- Ben-David D, Kordevani R, Keller N, Tal I, Marzel A, Gal-Mor O, et al. Outcome of carbapenem resistant *Klebsiella pneumoniae* bloodstream infections. *Clinical Microbiology and Infection* 2012; 18:54–60.
- Deshpande P, Rodrigues C, Shetty A, Kapadia F, Hegde A, Soman R. New Delhi Metallo-beta lactamase-1 in Enterobacteriaceae: Treatment options with carbapenem compromised. *J Assoc Physicians India* 2010; 58:147–149.
- Bhaskar MM, Anand R, Harish BN. Prevalence of blaNDM-Producing Blood Isolates of *Escherichia coli*, *Klebsiella* species and *Enterobacter* Species in a Tertiary Care Centre in South India. *J Micro Resear and Reviews* 2013; 6:61–68.
- Ulu AC, Kurtaran B, Inal AS, Kömür S, Kibar F, Çiçekdemir HY, et al. Risk Factors of Carbapenem-Resistant *Klebsiella pneumoniae* Infection: A Serious Threat in ICUs. *Medical Science Monitor: International Medical Journal of Experimental and Clinical Research* 2015; 21:219.
- Lin YT, Jeng YY, Chen TL, Fung CP. Bacteremic community-acquired pneumonia due to *Klebsiella pneumoniae*: clinical and microbiological characteristics in Taiwan, 2001–2008. *BMC Infect Dis* 2010; 10:307.
- Yao B, Xiao X, Wang F, Zhou L, Zhang X, Zhang J. Clinical and molecular characteristics of multi-clone carbapenem-resistant hypervirulent (hypermucoviscous) *Klebsiella pneumoniae* isolates in a tertiary hospital in Beijing, China. *Int J Infect Dis* 2015; 37:107–12.
- Zhang R, Lin D, Chan EW, Gu D, Chen GX, Chen S. Emergence of carbapenem-resistant serotype K1 hypervirulent *Klebsiella pneumoniae* strains in China. *Antimicrobial Agents and Chemotherapy* 2016; 60:709–11.
- Lee CH, Liu JW, Su LH, Chien CC, Li CC, Yang KD. Hypermucoviscosity associated with *Klebsiella pneumoniae*-mediated invasive syndrome: a prospective cross-sectional study in Taiwan. *International Journal of Infectious Diseases* 2010; 14:e688–92.
- Tan TY, Cheng Y, Ong M, Ng LS. Performance characteristics and clinical predictive value of the string test for detection of hepato-virulent *Klebsiella pneumoniae* isolated from blood cultures. *Diagnostic Microbiology and Infectious Disease* 2014; 78:127–8.
- Yu WL, Lee MF, Tang HJ, Chang MC, Chuang YC. Low prevalence of *rmpA* and high tendency of *rmpA* mutation correspond to low virulence of extended spectrum β-lactamase-producing *Klebsiella pneumoniae* isolates. *Virulence* 2015; 6:162–72.
- Yong D, Toleman MA, Giske CG, Cho HS, Sundman K, Lee K, et al. Characterization of a new metallo-β-lactamase gene, blaNDM-1, and a novel erythromycin esterase gene carried on a unique genetic structure in *Klebsiella pneumoniae* sequence type 14 from India. *Antimicrobial Agents and Chemotherapy* 2009; 53:5046–54.
- Oteo J, Domingo-García D, Fernandez-Romero S, Saez D, Guilo A, Cuevas O, et al. Abdominal abscess due to NDM-1-producing *Klebsiella pneumoniae* in Spain. *Journal of Medical Microbiology* 2012; 61:864–7.
- Shankar C, Nabarro LE, Ragupathi NK, Sethuvel DP, Daniel JL, Veeraraghavan B. Draft genome sequences of three hypervirulent carbapenem-resistant *Klebsiella pneumoniae* isolates from bacteremia. *Genome Announcements* 2016; 4:e01081–16.
- Zhang Y, Zeng J, Liu W, Zhao F, Hu Z, Zhao C, Wang Q, Wang X, Chen H, Li H, et al. Emergence of a hypervirulent carbapenem-resistant *Klebsiella pneumoniae* isolate from clinical infections in China. *J Infect* 2015; 71:553–60.